IMMUNOLOGICAL STUDIES OF COMBINED FOOT AND MOUTH DISEASE AND RIFT VALLEY FEVER OIL VACCINE

Walaa ,S. Shabana^a;Bakr, A. Abdalla^a; Eman, M. ELShalami^a; I.M. Reda^b; M. A. Shalaby^b

^aVeterinary Serum and Vaccines Research Institute, Abbassia, Cairo, Egypt. ^bCairo University, Faculty of Vet. Med., Dept. of Virology.

Abstract

Foot-and-mouth disease (FMD) is an economically important disease of cloven-hoofed animals. In Egypt, FMD assumes as enzootic form and attacks susceptible animals causing high losses in milk and meat production. Rift valley fever virus causes serious and fatal disease in animals and man. It produces high abortion rate among pregnant ewes and cows, causes heavy mortalities in young lambs and calves. In the present study the Montanide ISA 50 oil adjuvanted combined FMD/RVF vaccine was tested in sheep and compares it with single vaccines either FMD or RFV alone. The mean of antibody titers continued with the protective level till the 32 to 36th week post vaccination, In single vaccination either FMD or RFV alone while till to 40th WPV in combined FMD/RVF vaccines improve the immune response against FMD and RVF, giving high titer of antibodies against both diseases, and long duration of immunity in combined FMD/RVF vaccines.

Keywords : Foot and mouth diseasevaccine, Rift valley fever vaccine, Combined vaccine, Montanide ISA 50.

1. INTRODUCTION

Foot and mouth disease is one of the most troubles worlds wide viral disease of animals specially cloven footed of both wild and domestic animals (1). The causative agent is a single stranded positive- sense RNA virus that belongs to the genus Aphthovirus in the family Picornaviridae. There seven are immunologicaly distinct serotype of FMD virus, namely, O, A, C, Asia1, Sat1, Sat2 and Sat3 (2). In Egypt, the disease is enzootic and outbreaks have been reported since 1950 (3). FMD serotype O was the most prevalent since 1960 and onwards (4-6).Since 1950,1953 and 1956 serotype A didn't recorded in Egypt (4), serotype A FMD virus introduced to Egypt through live animals importation, and the sever clinical signs occurred among cattle and buffaloes (7). Recently FMD serotype SAT 2 outbreaks in Egypt were reported in eight (8) out of 27 governorates concentrated mainly in the Delta area and very few along the Nile in the southern parts of the country (8-9). The control of FMD in animals was considered to be important to effectively contain the disease in endemic areas, so that vaccination of animals is effective in limiting the spread of FMD (**10**).Most foot-and-mouth disease vaccines are made BEI (binary of Ethyleneinmine) inactivated virus that is adjuvanted with either aluminum hydroxidesaponin (AS) or oil adjuvant. Oil adjuvants are generally preferred over AS vaccines because among other advantages, they produce longer lasting immunity (11).

Rift Valley Fever (RVF) is an acute, sub acute or mild arthropod born viral disease of many species of animals as well as human being. The disease characterized by high mortality rates among calves and lambs as well as abortion of pregnant ewes and cows (13). RVF causes serious and fatal disease in animals and human being characterized by a short incubation period, fever, leucopenia and necrotic changes in the liver (14). RVFV is a negative-strand RNA virus belongs to the family Bunyaviridae, genus Phlebovirus (15&16). RVFV has traditionally caused recurrent outbreaks affecting humans and ruminants predominantly in Sub-Saharan Africa, but spread to Egypt in 1977 and to the Arabian Peninsula in 2000(17). after the appearance of the disease, identification and isolation of virus occurred and the Egyptian authorities succeeded in preparing a safe potent inactivated vaccine (18). Other studies were conducted by (19) to improve the vaccine and to raise its efficiency.

Due to the danger of both FMD and RVF diseases, systemic vaccination and quarantine measures are usually applied specially in enzootic areas as effective control measures. Combined vaccines are used for many human and animal diseases. However we have very few examples of combinations comprising anti-FMD valiancy which would allow easy immunization without additional handling.

Many authors recommended the use of combined vaccines against some infectious diseases in cattle and sheep that revealed good immunity as single vaccine (20).

The increase number of vaccines which are administered to the animal at different age and time make it necessary to study the immune response of animals vaccinated with two vaccines at the same time as a combined vaccine and compare it with the single vaccine to save efforts and times at launching vaccination campaign for more than one disease.

2. Material and Methods

2.1.AnimalsTwenty one adult susceptible sheep local breed of about 35-50 kg body weight, clinically healthy and free from antibodies against FMD and RVF viruses before the experimental work using serum neutralization test according to (21).

2.2.FMD viruses

The viruses usedin this study were locally isolated FMDV strains $O_1/3/93$, A/1/ Egypt 2006 and SAT2/2012 of cattle origin . The viruses were typed at Veterinary Serum and Vaccine Research Institute, Abbasia, Cairo and confirmed by Pirbright, World Reference Laboratories, United Kingdom.The antigens are stored at -70°C and used for preparation of vaccines and serologically tested for determination of antibodies against FMD virus types.

2.3.RVF virus (ZH 501)

The original virus was that isolated from a human patient in Zagazig, Sharquia province during 1977 outbreak and supplied by NAMRU-3 after being identified to be RVF virus. It was twice passaged I/C into suckling mice and has a final titer of 2×10^7 MICLD₅₀/ml. It was considered as the seed virus and preserved at -70°C.

3. Titration of FMD and RVF antigensused in vaccines preparation

3.1.Titration of FMD virus in tissue culture plates to detect the infectivity titer which expressed as log_{10} TCID₅₀ as described by (22)

3.2. titration of RVF virus in tissue culture tubes as recommended by (17&18) for detection of infectivity titer which expressed as \log_{10} TCID₅₀ as described by (22)

Complement fixation test used for detection of antigenicity of both FMD and RVF viruses used in vaccines preparation.

4. Virus inactivation and safety testing

FMD virus strains O₁ /Aga/ 93 , A/1/Egypt /2006 and SAT 2/2012 were inactivated by 0.1% M Binary ethylene amine (BEA; Sigma) as previously described (*23*).

Montanoid Oil : ISA 50 Montanoid Oil was obtained from Seppic, Paris, France.

5.Preparation of the oil adjuvant vaccines

The inactivated and clarified virus harvest was concentrated with 8% (w : v) polyethylene glycol

(PEG-6000) The inactivated oil adjuvenated FMD Vaccines were formulated according to (22). The ratio of the aqueous antigen to the oil adjuvant was 50:50 . The emulsions were produced by recycling the aqueous antigen-oil mixture several times. Sterility and safety of the prepared vaccines were done according to (23).

6.Preparation of combined oil adjuvant vaccine (FMD and RVF viruses)

The combined vaccine prepared from the previous inactivated FMD and RVF viruses is prepared as follows:

Mixing (4) parts of inactivated FMD virus with (1) part of inactivated RVF virus.That aqueous antigen mixture added in equal volumes (v/v) to (oil phase emulsion pH adjusted to 7.3-7.4 and mixed thoroughly.

7. Experimental Design

2. RESULTS

The obtained results are shown in tables (1-2&3) . It revealed that serum antibody protective titer against FMDV evaluated by mean of SNT started at 3^{rd} week post vaccination with FMD vaccine with the titer of(1.68-1.65&1.63) log₁₀ fortypes O –A &SAT2 respectively ,while in combined vaccine the protective level at 2^{nd} WPV with the titer (1.6 - 1.59 &1.74) log₁₀ and (1.89,1.83&1.74) at 3^{rd} WPV fortypes (O -A &SAT2) respectively.

The higher antibody level following vaccination was at the 10^{th} week with the titer of (2.7 - 3.0 log₁₀) forFMD O,A &SAT 2 and combined respectively, The mean of antibody titers continued with the protective level till the 36^{th} WPV for (O, A) and 32^{th} WPV for type SAT2 , while in combined vaccine protective level continued till 40WPV for type (O, A) and till 36^{th} WPV for type

Three groups(five sheep for each) werevaccinated with the tested vaccines.First sheep group was injected with FMD vaccine, second group vaccinated with combined (FMD&RVF) vaccine, third group vaccinated with RVF vaccine and fourth group was kept as negative control non vaccinated two sheep for each group. Serum samples were collected weekly post vaccination for one month then every weeks post-vaccination till the protective antibody level declined to non protective level. The immune response was evaluated through the estimation of humoral immune level using SNT.

48

8.Serum neutralization test (SNT)

It was performed using the technique as described by (20).

9.Enzyme linked immunosrobent assay (ELISA) It was carried out according to the method described by (24).

SAT2 after that the immunity duration started to decline under the protective level by SNT.

From tables (4) we noticed that serum antibody protective titer against RVF evaluated by mean of SNT started at 2nd week post vaccination with FMD vaccine with the titer of (1. 8 & 2.07) \log_{10} forsheep vaccinated with RVF vaccine and combined vaccine respectively. The higher antibody level following vaccination was at the 10th week with the titer of (3.1-3.28) \log_{10} forsheep vaccinated with RVF vaccine and combined vaccine respectively. The mean of antibody titers continued with the protective level till 36th week post vaccination in sheep the vaccinated with RVF vaccine and till 40th WPV in combined vaccine then started to decline under the protective level.

Weeks	Antib	ody titre	agains	t FMD	V type		An	tibody i	titre ag	ainst F	MDV	
post		O_1 in	FMD ve	accine		Mea	typ	-	n com		FMD	Mea
vaccina						n			RVF va			n
tion	1	2	3	4	5		1	2	3	4	5	
0	0	0.3	0.45	0	0.15	0.18	0.0	0.3	0.45	0.0	0.15	0.18
1	0.9	0.9	0.6	0.45	0.6	0.7	0.3	0.6	0.9	0.3	0.9	0.6
2	1.05	1.2	1.2	1.05	1.2	1.14	1.5	1.5	1.65	1.8	1.65	1.6
3	1.55	1.65	1.65	1.8	1.8	1.69	1.8	1.65	1.8	2.1	2.1	1.89
4	1.65	1.8	1.8	2.1	1.95	1.86	1.9 5	2.1	2.25	2.4	2.7	2.28
6	2.25	2.4	2.25	2.4	2.4	2.34	2.5 5	2.7	2.7	2.7	2.85	2.7
8	2.25	2,7	2.4	2.7	2.7	2.55	2.5 5	3.0	2.85	3.0	3.0	2.9
10	2.55	2.4	2.85	2.85	2.85	2.7	2.8 5	2.85	3.0	3.15	3.3	3.0
12	2.55	2.4	2.7	2.85	2.85	2.67	2.8 5	2.85	3.15	3.3	3.3	3.09
14	2.55	2.4	2.55	2.85	3.15	2.7	2.7	2.7	3.0	3.3	3.45	3.03
16	2.4	2.25	2.55	2.7	3.0	2.85	2.7	2.7	3.0	3.3	3.45	3.03
18	2.4	2.25	2.4	2.7	3.0	2.55	2.7	2.7	3.0	3.15	3.45	3.0
20	2.4	2.1	2.4	2.7	2.7	2.46	2.4	2.55	2.85	3.0	3.15	2.8
22	2.25	2.1	2.1	2.7	2.7	2.37	2.4	2.55	2.85	3.0	3.0	2.76
24	1.95	1.95	1.8	2.4	2.55	2.13	2.2 5	2.4	2.55	2.7	2.7	2.5
28	1.95	1.8	1.65	2.1	2.4	1.98	2.2 5	2.1	2.25	2.7	2.55	2.37
32	1.8	1.65	1.5	1.95	2.4	1.8	2.1	1.8	2.15	2.4	2.4	2.2
36	1.5	1.5	1.35	1.65	2.1	1.62	1.9 5	1.8	1.95	1.95	2.4	2.0
40	1.2	1.35	1.2	1.35	1.65	1.35	1.6	1.5	1.5	1.65	2.1	1.68
44	0.9	0.9	0.75	1.05	0.96	1.02	5 1.5	1.2	1.2	1.5	1.8	1.44

Table (1) Frequency of serum antibody titers against type (O), in sheep vaccinated with trivalent FMD vaccine and combined FMD &RVF vaccines by using SNT expressed log_{10}

*Antibody titers expressed as log₁₀ TCID₅₀

50

		Antibody titre against FMDV type A in FMD vaccineAntibody titre against FMDV type A											
Weeks	D vacci	ne	Me	type A									
post						an	in	VF	n				
vaccinati						_			vaccin	e		-	
on	1	2	3	4	5		1	2	3	4	5		
0	0.3	0.3	0.4 5	0.15	0.15	0.3	0.3	0.3	0.45	0.15	0.15	0.27	
1	0.8	0.8	0.5	0.35	0.5	0.8	0.3	0.6	0.9	0.3	0.9	0.6	
2	1.05	1.2	1.2	1.2	1.2	1.0 5	1.5	1.5	1.65	1.65	1.65	1.59	
3	1.65	1.6 5	1.6 5	1.65	1.65	1.6 5	1.8	1.8	1.8	1.95	1.8	1.83	
4	1.8	1.6 5	1.6 5	1.8	1.65	1.8	2.1	1.95	1.95	2.1	1.95	2.0	
6	2.1	1.8	2.1	2.1	1.8	2.1	2.4	2.4	2.4	2.7	2.4	2.5	
8	2.25	2.1	2.4	2.4	2.4	2.2 5	2.55	2.55	2.7	2.7	2.7	2.64	
10	2.7	2.5 5	2.7	2.85	2.7	2.7	3.0	3.0	3.15	3.0	3.15	3.0	
12	2.55	2.4	2.7	2.85	2.7	2.5 5	3.0	3.0	3.3	3.3	3.15	3.15	
14	2.4	2.4	2.5 5	2.7	2.55	2.4	2.7	2.7	3.0	3.0	3.0	2.88	
16	2.4	2,4	2.5 5	2.55	2.55	2.4	2.7	2.7	3.0	3.0	3.0	2.88	
18	2.4	2.2 5	2.4	2.4	2.4	2.4	2.7	2.55	2.7	2.85	2.7	2.7	
20	1.95	2.2 5	2.4	2.4	2.4	1.9 5	2.4	2.25	2.55	2.7	2.55	2.5	
22	1.95	2.1	2.1	2.25	2.25	1.9 5	2.1	2.25	2.4	2.4	2.4	2.3	
24	1.8	1.8	2.1	2.25	2.25	1.8	1.95	2.1	2.4	2.25	2.25	2.2	
28	1.5	1.5	1.8	1.95	2.1	1.5	1.8	1.8	2.1	2.25	2.25	2.0	
32	1.5	1.5	1.5	1.95	1.8	1.5	1.65	1.8	1.8	1.95	2.1	1.86	
36	1.2	1.3 5	1.2	1.8	1.65	1.2	1.5	1.5	1.65	1.8	1.95	1.68	
40	0.9	1.2	0.9	1.5	1.35	0.9	1.5	1.2	1.5	1.5	1.8	1.5	
44	0.6	0.9	0.7	2.1	1.05	0.6	1.2	0.9	1.2	1.2	1.5	1.2	
			5										

Table (2) Frequency of serum antibody titers against type (A), in sheep vaccinated with trivalent FMD vaccine and combined FMD &RVF vaccines by using SNT expressed log₁₀

*Antibody titers expressed as log₁₀ TCID₅₀

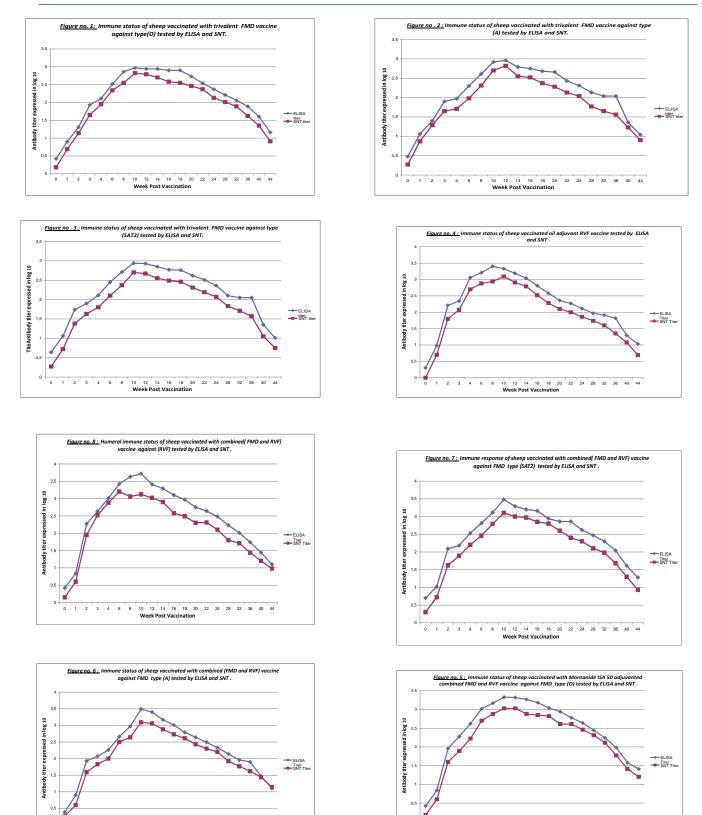
	Antib	ody titi	re aga	inst FA	MDV		Antil					
Weeks	typ	oe A in	FML) vaccii	ne	Mea		i			Mea	
post						n	in c	VF	п			
vaccinatio												
n	1	2	3	4	5		1	2	3	4	5	
0	0.3	0.3	0.4 5	0.15	0.15	0.27	0.3	0.3	0.4 5	0.15	0.15	0.3
1	0.6	0.6	0.6	0.9	0.9	0.72	0.6	0.6	0.6	0.9	0.9	0.72
2	1.2	1.2	1.2	1.5	1.8	1.38	1.5	1.65	1.6 5	1.8	2.1	1.74
3	1.65	1.5	1.5	1.7	1.8	1.63	1.8	1.8	2.1	1.95	2.1	1.95
4	1.8	1.65	1.6 5	1.8	2.1	1.8	2.1	2.1	2.2 5	2.1	2.4	2.2
6	2.1	1.8	2.1	2.1	2.4	2.1	2.4	2.25	2.5 5	2.4	2.7	2.5
8	2.25	2.1	2.4	2.4	2.7	2.37	2.55	2.7	2.8 5	2.85	3.0	2.8
10	2.7	2.4	2.7	2.85	2.85	2.7	3.0	2.85	3.1 5	3.15	3.3	3.1
12	2.55	2.55	2.7	2.7	2.85	2.67	3.0	3.0	3.0	3.0	3.0	3 .0
14	2.4	2.55	2.5 5	2.55	2.7	2.55	2.85	3.0	3.0	3.0	3.0	2.97
16	2.4	2.25	2.5 5	2.55	2.7	2.49	2.7	2.85	3.0	2.85	2.85	2.85
18	2.25	2.55	2.4	2.55	2.55	2.46	2.7	2.85	2.8 5	2.7	2.85	2.8
20	1.95	2.25	2.4	2.4	2.55	2.31	2.25	2.7	2.8 5	2.4	2.7	2.6
22	1.95	2.1	2.1	2.25	2.55	2.19	2.25	2.4	2.5 5	2.25	2.55	2.4
24	1.8	1.8	2.1	2.25	2.4	2.07	2.1	2.25	2.4	2.25	2.4	2.3
28	1.8	1.65		1.8		1.89		2.1	2.1	2.1	2.25	2.1
32	1.65	1.5	1.6 5	1.95	1.8	1.71	1.8	1.95	1.9 5	1.95	2.25	1.9
36	1.5	1.2	1.3 5	1.65	1.65	1.47	1.5	1.65	1.6 5	1.8	1.8	1.68
40	0.9	0.75	0.9	1.2	1.5	0.9	0.9	1.2	1.3 5	1.5	1.5	1.3
44	0.6	0.6	0.6	0.9	1.05	0.75	0.6	0.9	0.9	1.05	1.2	0.96

Table (3) Frequency of serum antibody titers against type (SAT 2), in sheep vaccinated with trivalent FMD vaccine and combined FMD & RVF vaccines by using SNT expressed log₁₀

52

	Antib	ody titr	e aga	inst R	VF of		Anti	RVF				
Weeks	shee	p vacci	inated	l with l	RVF	Mea	J I					
post						п	combined FMD &RVF					
vaccinati					_			vaccine				
on	1	2	3	4	5		1	2	3	4	5	
0	0.0	0.0	0.0	0.0	0.0	0.0	0.3	0.0	0.3	0.0	0.15	
1	0.6	0.6	0.9	0.6	0.6	0.7	0.3	0.6	0.9	0.3	0.9	
2	1.2	1.8	2.1	1.95	1.8	1.8	1.8	1.8	2.4	2.4	1.95	2
3	1.8	2.4	2.4	2.7	2.4	2.34	2.4	2.4	2.7	2.7	2.4	2
4	2.4	2.7	2.8 5	2.85	2.7	2.7	2.7	2.7	3.15	3.0	2.85	2
6	2.7	3.0	3.0	2.85	2.85	2.88	3.15	3.1 5	3.3	3.3	3.15	
8	2.7	3.0	3.0	3.0	3.0	2.94	3.1	2.7	3.3	3.3	3.3	
10	3.0	3.0	3.3	3.15	3.0	3.1	3.1	3.3	3.6	3.15	3.1	
12	2.85	2.85	2.8 5	3.0	3.0	2.9	3.0	2.8 5	3.3	3.0	2.85	
14	2.7	2.7	2.8 5	2.85	2.85	2.79	3.0	2.7	3.15	3.0	2.55	
16	2.55	2.4	2.7	2.55	2.4	2.52	2.7	2.4	2.85	2.7	2.25	2
18	2.4	2.1	2.4	2.1	2.4	2.28	2.7	2.4	2.7	2.7	1.95	2
20	2.1	1.8	2.4	1.95	2.1	2.1	2.4	2.1	2.7	2.7	1.8	
22	2.1	1.8	2.2 5	1.95	1.95	2.0	2.25	2.4	2.7	2.4	1.8	2
24	1.8	1.65	2.2 5	1.8	1.8	1.86	2.4	2.1	2.4	2.1	1.5	
28	1.5	1.5	2.1	1.8	1.8	1.7	1.8	1.8	2.1	1.8	1.5	
32	1.5	1.5	1.8	1.8	1.5	1.6	1.8	1.8	2.1	1.8	1.5	
36	1.35	1.35	1.5	1.35	1.2	1.35	1.5	1.6 5	1.65	1.8	1.8	
40	1.2	0.9	1.3 5	1.05	0.9	1.08	0.9	1.2	1.35	1.5	1.5	
44	0.6	0.6	0.9	0.6	0.75	0.69	0.9	1.0 5	1.05	0.9	1.2	

Table (4)Frequency of serum antibody titers against RVF in sheep vaccinated with RVF
and combined FMD &RVF vaccines by using SNT expressed log10





0 1 2 3 4 6 8 10 12 14 16 18 20 22 24 28 32 36 40 44 Week Post Vaccination 0 1 2 3 4 6 8 10 12 14 16 18 20 22 24 28 32 36 40 44 Week Post Vaccination

4. DISCUSSION

Foot and mouth disease is considered the major infectious disease affecting cattle, buffaloes, sheep and other cloven footed animals. It characterized by fever, vesicular eruptions of the mouth mucosa, teats and the coronary bands of the hooves. The disease in sheep tends to be mild, transient or even inapparent infection, while in lambs it may be peracute and cause sudden death. It is enzootic in Africa, Asia, Europe, Philippines and South America (25). The control of FMD in sheep was considered to be important to effectively contain the disease in endemic areas, (10).

Rift valley fever virus causes serious and fatal disease in animals and man. It produces high abortion rate among pregnant ewes and cows, causes heavy mortalities in young lambs and calves (26).

The progress in vaccine production is directed towards the selection of the proper adjuvant that can elaborate high and long lasting immunity. Adjuvants can influence the immune response and prolong the immune response and stimulate specific components of the immune response either humoral or cell mediated immunity (27).

Three experimental of FMD, RVF and combined FMD/RVF vaccines batches. The viruses inactivated by binary ethyleimimine and adjuvanted with Montanide ISA50 oil adjuvant.

Regarding the results of innocuity, sterility and potency of different inactivated prepared vaccines, the results obtained in there is no detection of cytopathic effect (CPE) on BHK cells after three passages of the inactivated viruses used in vaccine preparation, indicating that there is no viable viral residues after the inactivation process. Also, FMD lesions did not appear on susceptible sheep when inoculated at different sites of the tongue.

in tables (2-3&4) findings agreed with (28)

who found that the peak titer with oil FMD vaccines was not reached before 60-80 days post vaccination. Our results also agreed with (29) who recorded that the immunogenicity of FMD vaccines can be considerably increased by the use of proper adjuvants, in FMD vaccines prepared for cattle and sheep.

54

Our results also agreed with (29) who recorded that the immunogenicity of FMD vaccines can be considerably increased by the use of proper adjuvants, in FMD vaccines prepared for cattle and sheep. Also our results were supported by (30) who used the serum neutralizing antibody assay for determining the potency of FMD vaccines The finding indicated that protective capacity of the prepared vaccine.

The obtained results were in agreement with (31) who found that oil FMD vaccines gave high and long duration immunity, while disagreed with *Samir* (32 &33) in that the protective titer of antibody continued with protective level till the 38^{th} week post vaccination, then decreased than that results obtained by him in which the protective level started at 3^{rd} week post vaccination.

These results were supported by (34) who compared between using of different Montanide ISA oil adjuvants and different Montanide IMS oil adjuvants in emergency FMD vaccine for Guinea pigs. Regarding the study of humoral immune response of sheep vaccinated with combined FMD and RVF vaccine, the obtained results of SNT.

The obtained results in **tables** (4) agreed with those of (35&36) where they mentioned that the use of oil adjuvant in FMD vaccine involved a more efficient antigen stimulus and more sustained antibody response, (37)who found that mixed vaccination with anthrax and FMD were as good as FMD vaccination on its own, (38) mentioned that the antibodies developed from vaccination of cattle by FMD virus, rabies and *Brucella*

abortus were as high as of the individual vaccine of each,

Our results were agreed with (39) who prepared RVF/FMD combined vaccine which protect animals well against challenge with the virulent viruses, (40&41) who studied the vaccination of sheep with combined RVF and sheep pox vaccine which protect animals well against challenge with the virulent viruses, Assessment of neutralizing antibodies of RVF after vaccination was considered by (42) as a way for evaluating the protective capacity of prepared vaccines. (43) suggested that the protective titer was 1.7 log₁₀, while (44&45)mentioned that serum neutralizing protecting titer considered not less than 1.0 log₁₀. These results agreed with (46).

The protective antibody level of sheep vaccinated with Montanide ISA 50 as an oil adjuvant (FMD and RVF) vaccine results revealed that the prepared vaccine improve the immune response against FMD and RVF, giving high titer of antibodies against both diseases and indicated that the combined FMD/RVF vaccine conferred long duration of immunity than conferred by single vaccine against these diseases when used alone.

Finally we can concluded that using of combined FMD/RVF vaccines give long duration of immunity and must be used to save efforts and times at launching vaccination.

5. REFRANCES

1.Radostits, O.M.; Blood, D.C. and Goy, C.C.
1995.Veterinary Medicine, P. 965973.Educational low priced blooks scheme,
Funded by the British Government, 8th Ed
2.Belsham, G.J. 1993. Distinctive features of
FMDV, a member of the Picorna virus family,
aspects of virus protein synthesis, protein
processing and structure. Progress in
Biophysics and Molecular Biology 60: 241-260.

3. Moussa, A.A.M.; Stouraitis, F.; Ibrahim, M.H.; Daoud, A. and Hussein, K. 1974. Foot and Mouth Disease vaccine production in baby hamster kidney (BHK21) cells in suspension in Egypt.Bull. Off. Int. Epiz., 81 (11-12): 1043-1054.

4.Zahran,G.E.D. 1960. "Foot and mouth disease in southern region of URA".

5. Daoud, A., Omar, A., El Bakry, M., Metwally, N., El Mekkawi, M. and El Kilany, S. 1988.Strains of Foot and Mouth Disease virus recovered from 1987 outbreak in Egypt. J. Egypt. Vet. Med. Ass., 48 (1): 63-71.

6.Farag. M. A., A.M.A.Aggour and A.M.Daoud .2005."Elisa as rapid method fordetecting the correlation between the field isolates of FMD virus and the current used vaccine strain in Egypt". Cairo. Vet. Med. Journal, volume <u>53</u>, No1, Jan. 2005.

7.Abd El-Rahman A.O.,El-Kilany S.,Farag M.A.,El-Garf E.,Abu Elyazid M. And zidan S.M. 2006." Isolation and identification of FMD virus during an outbreak of 2006 in Egypt." Kafr El-Sheikh Vet.Med.J. Egypt ,<u>4</u> (1):452-463.

8.Lockhart, C.; Sumption, K.; Pino, J. and Lubroth, J. 2012.Foot-and-mouth disease caused by serotype SAT2 in Egypt and Libya.Empress watch, 25 : 1-7

9.Shawky M., Abd El-Aty M., Hiam. M.Fakhry,, Hind M.Daoud, Ehab El-Sayed I., Wael Mossad G.,Sonia A.Rizk, Abu-Elnaga H., Mohamed A.A., Abd El-kreem A. and Farouk E.M. : Isolation and Molecular Characterizationof Foot and Mouth Disease ,SAT2 Virus during Outbreak 2012 in Egypt . J.Vet. Adv, 3(2): 60-68 (2013).

10. Nair,S.P. and Sen,A.K. 1992. "A comparative study on the immune response of sheep to FMD virus vaccine type Asia1 prepared with different inactivators and adjuvants". Comp. Immunol. Microbiol. Infect. Dis., 15(2): 117-124.

11.Doel, T.R. 2003 .Review of FMD vaccines.Virus Research, 91: 81-99.

12.Easterday,B.C.; Murphy,L.C. and Bennett,D.G. (1962): "Experimental RVF in lambs and sheep". Amer.Vet.Res., 23: 1231-1240.

13.Daubney,R.; Hudson,J.R. and Graham,P.C.1931. "Rift valley fever or enzootic hepatitis as undescribed diseases of sheep, cattle and man from East Africa". J.Path.Bact., 34 (1): 545-579.

14. Ikegami T., Peters C.J. and Makino S. 2005. Rift Valley Fever virus nonstructural protein NSs promotes viral RNA replication and transcription in a minigenome system. J.Virol, 2005 May(b); 79(9) :5606-15

15.HanafiA.H. David , MagdyD.Saeid,Atef Soliman,Eman Medhat ,Adel dasset,Zaid,DanialE.Szumumlas,Tennets C. Earhart .2011. Virus isolation and hight population in delicate culex antenna ,diptra culetin vector of Rift Valley Fever virus during outbreak in Nile Delta of Egypt Actatopia xxx:xxx-xxx

16.Mandell R. B., Ramesh Koukuntla, Laura J.K. Mogler, Andrea K. Carzoli, Alexander N. Freiberg, Michael R. Holbrook, Brian K. Martin, William R. Staplin, Nicholas N. Vahanian, Charles J. Link, Ramon Flick. 2010. A replication-incompetent Rift Valley fever vaccine: Chimeric virus-like particles protect mice and rats against lethal challenge. Virology 397 (2010)187-19814 17.El-Nimr,M.M. 1980. "Studies on the inactivated vaccine against Rift Valley fever". Ph.D.Thesis (Microbiology), Fac.Vet.Med., Cairo University.

18.Taha,M.M. 1982. "Studies on inactivated vaccine of RVF virus". Ph.D.Thesis (Microbiology), Fac.Vet.Med., Cairo University.

19. Ebied,M.H.; M.Roukaya Ossman; A.Z.Hussein and R.A.Diab.1999.

"A multicomponent vaccine against blackleg, haemorrhagic septicaemia, FMD and EVF". Egypt.J.Agric.Res., 77 (4): 1877-1885.

56

20. Ferreira, M E .1976. Prubade microneutralization proestuase de anticurpos de la fiebre aftose.Blth. Centropan Americano Fieber Aftosa, 21 and 22: 17-24.

21. Reed LJ and Muench H A .1938. simple method for estimating fifty percent end points. Am J Hyg.; 27:493–8.

22.Barnett, P., Pullen, L. Williams, L. and Doel, T.R. 1996. International bank for footand-mouth disease vaccine: assessment of Montanide ISA 25 and ISA 206, two commercially available oil adjuvants. Vaccine 1996;14(13):1187–1198.

23.OIE Manual .2000 . OIE manual of standards : Anon. Section 2.1. List - Diseases, Chapter 2.1.1. Foot-and-mouth disease. OIE manual of standards for diagnostic tests and vaccines; 4th Ed. 2000, Paris, 77-92

24.Voller A , Bid Well D and Bartleha.1976. Micro plate enzyme immuno assay for the immuno diagnosis of virus infection . Am. Soc. For Micro.(506-512) .

25.Blood,D.C.; Radostitis,O.M. and Henderson,J.A.C. 1995."Veterinary medicine". A textbook of the diseases of cattle, sheep, goats and horses. Eighth edition, ELBS and Bailliere Tindall.

26.Wood,O.L.; Meegan,J.M.; Morrill,J.C. and Stephenson,E.H.1990. "Virus infections of ruminants". Edited by Diner,Z. and Morein,B., 481-493. Amestrdam, Netherlands, Elsvier Science Publishers B.V.

27.Dalsgarrd,K.; Hilgers,L. and Trouve,G.1990. "Classical and new approaches to adjuvant use in domestic food animals". Adv.Vet.Sci.Comp.Med., 35: 121-159.

28. Graves, J.H.; Cowan, K.M. and Trautman, R. 1968. Immunochemical studies of FMDV characterization of RNA free virus like particles.Virology, 34: 269-274.

- 29.Solymon,F. and Czelleng .1977. Studies on the correct quantitative relation of antigen components in one and trivalent FMD vaccine preparations". Int.Symp. on FMD, Lyon, Develop.BioI.Stand, Vol. 35, pp.289- 294.
- 30. Hingley, P.J. and Pay, T.W.F. 1987. "Sources of variability in FMD vaccine potency estimates based on serum neutralizing antibody assay". J.Biol.Stand., 15: 127-142.
- 31.Barteling,S.J. and Vreeswij,K.J. 1991. "Development in foot and mouth disease vaccine". Vaccine, 9(2): 75-88.
- 32.Samir,M.A.A.2002. "Studies on preparation of newly oil adjuvanted FMD vaccine". Ph.D.Thesis (Virology), Fac.Vet.Med., Cairo University.
- 33.Hiam M Fakhry, Sonia A Rizk, Hany I Abu-Elnaga,Wafaa Deghaidy, Abeer A Talaat and Hegazi A Z 20412.Field Application Of Bivalent Foot And Mouth Disease Vaccine Adjuvanted With Montanide ISA (25, 50, 206, 1113) And IMS 3015 as An Alternative To Aluminum Hydroxide Gel . Zag. Vet. J. (ISSN. 1110-1458) Vol. 40 No. 5 (2012) pp.188-195
- 34.Barnett,P.V.; Pullen,L.; Warder,P. and Stathen,R. 1999. "International bank for foot and mouth disease vaccine (preliminary studies on emergency foot and mouth disease vaccines formulated with montanide IMS -Immunosol-, a new concept in oil adjuvancy", European Commission for the Control of FMD, Aldershot, United Kingdom, 14-18 September 1998, Appendix 37: 268-271.
- 35. Cunliffe,R. and Graves,J.H.1963. "Formalin treated FMDV comparison of two adjuvants in cattle". J.Comp.Vet.Sci., 27: 193-197.
- 36.Rivenson,S.; Sadir,A.M.; Caggino,O.P.; Zabal,O.P. and Laporte,O. 1982."Comparison of two foot and mouth disease vaccines (oil

emulsion and hydroxyl saponin) in cattle". Rev.Med.Vet., Argentina, 65: 364-370.

37.Nobili,I. and Colonna,V.1973. "Immunological response in sheep vaccinated against anthrax and FMD as represented by neutralizing antiviral antibodies". Veterinaria Italiana, 34: 117-126.

38. Favre,H.; Valette,L.; Precausta,P.; Roulet,C.; Brun,A.; Terre,J. and Stellmann,C.1976."Anti-FMD vaccines combined with other bacterial or viral vaccines". Dev.Biol.Stand., (35): 409-428.

39. Gihan,K.M.1990. "Studies on RVF among animals in Egypt". Ph.D.Thesis (Infectious Diseases), Fac.Vet.Med., Zagazig University.

40.Timour,N.M.1992. "Attempts of preparing a compound RVF vaccine with sheep pox vaccine for sheep in Egypt". M.V.Sc. Thesis (Infectious Diseases), Fac.Vet.Med., Alexandria University.

41.Zaki,F.F.; Khirat,A.Elian; Hala,A.Fadl and Wassel,M.S.1999."Trial of production and evaluation of combined vaccine from RVF, Pasteurella haemolytica and Pasteurella multocida inactivated with binary in cattle". Egypt.J.Agri.Res.,

42. Weiss,K.E. and Serfontein,J.W.1955. "The stability of RVF virus a cited by Weiss 1957". Bull.Epiz.Dis. of Afr., 5: 431-458.

43.Randall,R.; Binn,L.N. and Harrison,V.R.1964."Immunization against RVF virus – Studies on the immunogenicity of lyophilized formalin inactivated vaccine". J.Imm., 93 (2): 293-299.

44.Walker,J.S.; Stephen,E.L.; Remmel,N.S.; Carter,R.C.; Mitten,J.W.; Sohuli,L.G. and Klein,F. 1970. "The clinical aspects of RVF in house hold pets. II-Suscpetibility of the cat". J.Inf.Dis., 121: 19-24.

45.Pini,A.; Lund,L.J. and Davis,S.J. 197. "Fluorescent and neutralizing antibody response to infection by RVF virus". J.S.Afri.Z.Med.Ass., 44 (11): 161-165.

46.Gihan, K. M. and Khirate Abdel Megid.1997.Comparative studies on

IMMUNOLOGICAL STUDIES OF COMBINED FOOT AND MOUTH DISEASE AND RIFT VALLEY FEVER OIL VACCINE

serological response of locally produced live attenuated and inactivated Rift Valley vaccines. J. Egypt. Vet. Med. Ass. 57,No.1: 949-957.

دراسات مناعية على اللقاح المركب الزيتى لمرضى الحمى القلاعية وحمى الوادى المتصدع

ولاء شبانة شبانة عمر '، احمد عبدالله بكر '، المان محمد الشلقامي '، اسماعيل محمد رضا '، محمد عبد الحميد شلبي ' ' معهد بحوث الامصال واللقاحات البيطرية بالعباسية - قسم الحمي القلاعية ، 'كلية طب بيطري جامعة القاهرة -قسم الفير وسات الملخص العربي

نظرا لتعدد اللقاحات التى يتم التحصين بها فى أوقات و أعمار مختلفة للحيوانات فكان من الضرور در اسة تأثير التحصين بلقاحين مختلفين فى نفس الوقت ، لذا أجريت هذه الدر اسة لإنتاج لقاحمر كب لمرضى الحمى القلاعية ثلاثى العترة وحمى الوادى المتصدع كمحاولة لتحسين مناعة الحيوانات للقاح وذلك باستخدام مانتونيد أى أس أيه ٥٠ . بدأ مستوى الحماية المناعية للقاح الثلاثى للعترات الموجودة فى مصر (O A &SAT2) باستخدام مانتونيد أى أس أيه ٥٠ . بدأ مستوى الحماية المناعية للقاح الثلاثى للعترات الموجودة فى مصر (O A &SAT2) للعترات الموجودة فى مصر (O A &SAT2) باستخدام مانتونيد أى أس أيه ٥٠ . بدأ مستوى الحماية المناعية للقاح الثلاثى للعترات الموجودة فى مصر (O A &SAT2) لو ١٠ سجل اعلى مستوى مناعى فى الاسبوع الثالث للتحصين (٢,٧) للعترات الموجودة فى مصر (O A &SAT2) لو ١٠ سجل اعلى مستوى مناعى فى الاسبوع العاشر من التحصين (٢,٧) لو ١ و ١٠ سجل اعلى مستوى مناعى فى الاسبوع العاشر من التحصين (٢,٧) لو ١ و ١٠ سجل اعلى مستوى مناعى فى الاسبوع العاشر من التحصين والاسبوع الثالث للتحصين (٢,٧) بينما حقق اللقاح (O A (٥,١ - ١,٦ - ١,١) لو ١٠ سجل اعلى مستوى مناعى فى الاسبوع العاشر من التحصين (٢,٧) بو ١ و المن من الحمي التحمين والاسبوع التالية التحصين والاسبوع الاسبوع العاشر من التحصين والاسبوع القاح (O A) بينما حقق اللقاح (O A) مستوى مناعي أفضل حيث استمر حتى الأسبوع ٦٦ للعترة سات ٢ • ٤ بالنسبة لعترتى اللقاح (O A) و ١٠ سبوع ٢٦ للعترة سات ٢ وحتى الأسبوع ٦٦ من التحصين والأسبوع الثانى من (O A) بينما حقق اللقاح (A - O). كما أظهرت النتائج أن الأجسام المناعية وصلت لمستوى الحماية عند الأسبوع الثانى من والأسبو والنسبة للتحرين وكانت(١٨, - ٧, 1) لو ١٠ بالنسبة للقاح حمى الوادى المتصدع الأحادى والمركب مع الحمى القلاعية على التوالى واستمر المستوى المانيوي وكانت(المناعي حتى الأسبوع ٦٦ للعار ما المناعية وستمر مع الحمى القلاعية واستم والأسبوع الثاني ما واستمرى المستوى المانعي حتى الأمسوى النتائي من والأسبوي الما معال معنوى والأسبوع ٤٠ التومي حدى السبوع الثاني ما واستمر وكانت(١٨, - ٧, ١) لو ١٠ بالنسبة للقاح حمى الوادى المتصدع وحتى الأمادي مع الحمى القلاعية واستمر المستوى الماني عائمي ما مالي ما معال معال معال ما القاح والخاي ما اللقاح الأحمى ما القاح الأحمى ما القاح مامي مالمرك